


101

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Nucleotide Sequence of Cucurbit Aphid-Borne Yellow's Luteovirus

H. GUILLEY,¹ CATHERINE WIPF-SCHEIBEL,* K. RICHARDS, H. LECOQ,* AND G. JONARD

Institut de Biologie Moléculaire des Plantes du CNRS et de l'Université Louis Pasteur, 12 rue du Général Zimmer, 67084 Strasbourg Cédex, France; and *Station de Pathologie Végétale, INRA, Domaine Saint-Maurice, BP 94, 84143 Montfavet Cédex, France

Received February 2, 1994; accepted March 22, 1994

The nucleotide sequence (5669 residues) of the genomic RNA of cucurbit aphid-borne yellow's luteovirus (CABYV) is presented. Analysis of genome organization and sequence homologies indicate that CABYV is a member of luteovirus Subgroup 2 (other sequenced members: beet western yellow's virus, potato leafroll virus, and barley yellow dwarf virus, RPV isolate) and appears to be most closely related to beet western yellow's virus. © 1994 Academic Press, Inc.

Recent outbreaks of a severe yellowing disease of cultivated cucurbits in France have been attributed to a small isometric virus which is not mechanically transmitted, but which is efficiently transmitted in a persistent manner by the aphids *Myzus persicae* and *Aphis gossypii* (1). The virus, designated cucurbit aphid-borne yellow's virus (CABYV), is widespread in cucurbits throughout the Mediterranean basin and has also been detected in Asia and Africa (H. Lecoq, unpublished observations) and in North America (2). Serological analysis and nucleic acid hybridization experiments, along with virion morphology and the aforesaid vector transmission studies, indicated that CABYV is a member of the luteovirus group (1).

Sequence comparisons (3-5) have made it clear that luteoviruses can be divided into two major subgroups, which have been referred to (4) as Subgroup 1 (sequenced members: barley yellow dwarf virus (BYDV), PAV, and MAV isolates (6-7) and Subgroup 2 (sequenced members: beet western yellow's virus (BWYV (3)), potato leafroll virus (PLRV (8-10)) and BYDV, RPV isolate (11)). Recently, it has been suggested that soybean dwarf virus may belong to a third subgroup (12). Among the features distinguishing Subgroups 1 and 2 are the following: (i) Members of Subgroup 2 have a 5'-proximal open reading frame (ORF) encoding an approximately 29-kDa species, P1 (see Fig. 1A for ORF nomenclature), which is lacking in Subgroup 1, while the latter possess a short 3'-proximal ORF absent in the subgroup 2 members; (ii) the region of overlap between ORF 2 (P2) and ORF 3 in the Subgroup 2 viruses is much longer than in Subgroup 1 (Fig. 1A); (iii) the "core polymerase" domain on ORF 3 of the Subgroup 2 viruses displays

clear sequence affinity with the polymerase domain of southern bean mosaic virus while the same region of the Subgroup 1 viruses is related to the polymerase domain of the carmoviruses (see Ref. 5 for a review); (iv) there is considerable sequence homology in coding regions among the members of each subgroup except for ORF 1 of Subgroup 2. Intersubgroup sequence homology also exists but is less extensive, essentially being limited to the cluster of genes nearer the 3'-terminus, i.e. the coat protein (ORF 4), ORF 5, and ORF 6. Finally, it should be noted that, despite the aforesaid differences, the members of Subgroups 1 and 2 have certain features in common with respect to gene expression, including (a) the expression of ORF 3 as a ORF 2-3 fusion protein (P2/3) due to translational frameshift in the region of overlap between ORF 2 and ORF 3 (13-16), (b) expression of the cluster of three 3'-terminal genes from a single subgenomic RNA (17), and (c) expression of ORF 6 as an ORF 4-6 fusion protein (RT protein) due to translational read-through of the coat protein (ORF 4) termination codon (3, 16, 17, 18).

In this paper we describe the nucleotide sequence of 5669 residues for the genomic RNA from CABYV, obtained by characterization of cloned cDNA, and analyze its relatedness to other luteoviruses. Purified RNA from CABYV (isolate N) virions consists of a single RNA species of about 5600 nt, as judged by its mobility during electrophoresis under denaturing conditions in an agarose gel (data not shown). Sequence analysis of various luteoviruses has revealed highly conserved sequence domains in the coat protein cistron, making it possible to design luteovirus-specific primers as an aid to diagnosis and classification (19, 20). We have synthesized a 25-mer, 5'-ATTTCATGGTAGG(C/A)CTTGAG(T/C)ATTCC (oligo 1), complementary to residues 3784-3808 of the final CABYV sequence. This region encodes the amino

Sequence data from this article have been deposited with the EMBL Data Library under Accession No. X76931.

¹ To whom reprint requests should be addressed.

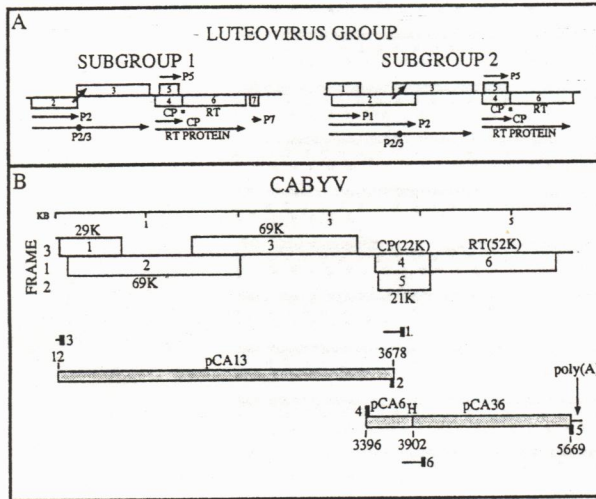


Fig. 1. Luteovirus genome organization. (A) Genetic organization of Subgroup 1 and Subgroup 2 luteoviruses. Open reading frames (ORFs) are indicated by hollow rectangles. ORFs in the map of the Subgroup 1 luteovirus (no ORF 1) have been numbered so that they correspond to the numbering of ORFs in Subgroup 2. The approximate site of frameshift between ORF 2 and ORF 3 is indicated by a slanted arrow and the approximate site of the resulting fusion between the ORF 2 and ORF 3 polypeptides (P2/3) is indicated by a black circle. The suppressible termination codon at the end of the coat protein (CP) cistron is indicated by an asterisk. RT, readthrough. (B) Position of long ORFs (hollow rectangles) in CABYV RNA (above) and the structure of cloned cDNA inserts (stippled rectangles) analyzed to determine the sequence (below). In the lower part of the figure, the numbered black squares (not to scale) represent some of the synthetic oligonucleotides used to prepare cDNA clones, to prime the polymerase chain reaction (PCR), or for direct sequence determination. In the latter case, the extent of sequence determined by primer extension is indicated by a horizontal line. The poly(A) tract enzymatically added to the 3'-end of the viral RNA and subsequently amplified by PCR using oligonucleotides 5 and 4 is represented by a thin line. H, *Hind*III.

acid sequence G(I/M)LK(A/S)YHE, which is perfectly or almost perfectly conserved in the coat protein of other Subgroup 2 viruses. Direct RNA sequence analysis using this primer to initiate reverse transcription (21) provided readable sequence of about 200 residues (Fig. 1B). For unknown reasons, attempts to produce and clone (22) long double-stranded cDNA fragments using oligo 1 to prime cDNA synthesis failed. However, use in the cloning protocol of another primer (oligo 2; complementary to residues 3655–3678) produced clones with suitably long cDNA inserts, one of which, pCA13 (Fig. 1B), was chosen for further analysis. The approximately 3700 residue cDNA insert of pCA13 was sequenced (23) on both strands principally by characterizing sets of 5'- and 3'-terminal nested deletions generated with a Pharmacia Double-Stranded Nested Deletion kit according to the supplier's instructions and whose extremities were then sequenced with the universal and reverse M13 primers. Some sequence data were also generated using synthetic primers designed to anneal to appropriate internal sequences in the insert.

To determine if the pCA13 insert extended to the 5'-terminus of the viral RNA, the sequence of the viral RNA was determined directly by reverse transcriptase-directed primer extension using oligo 3, which is complementary to residues 30–48 of the final RNA sequence (Fig. 1B). The resulting sequence was identical to that obtained by analysis of pCA13 up to a point 53 residues from the 5'-terminus of the cloned insert. At this point the two sequences diverged, with the sequence ladder obtained by primer extension ending with a strong-stop signal six residues upstream. The sequence determined by primer extension displays extensive homology with the 5'-terminal sequences reported for other luteoviruses (see below), providing additional evidence that the sequence obtained by primer extension corresponds to the authentic 5'-extremity. The presence of an "alternate" unrelated 5'-terminal sequence on pCA13 brings to mind observations with Scottish isolates of PLRV, in which the 5'-terminal 14 residues which displayed almost perfect sequence homology with an exon of tobacco chloroplast DNA ORF 196 (24). PCR experiments provided direct evidence for the existence of small amounts of host RNA–viral RNA chimeras in the viral RNA preparation, indicating that rare recombination events can occur between host and viral RNAs (24). The 53-residue "alternate" 5'-terminal sequence found in pCA13 did not display significant homology with sequences in databases but residues 1–36 were perfectly complementary to residues 4045–4080 from the CABYV coat protein cistron. Thus, in the case of pCA13, the alternate 5'-terminal sequence may be a cloning artifact.

Luteovirus RNAs are not 3'-polyadenylated. Consequently, CABYV RNA was treated with *Escherichia coli* poly(A) polymerase under conditions similar to those used to 3'-polyadenylate BWYV RNA (3) but when the resulting 3'-polyadenylated RNA was used as template for cDNA synthesis and cloning by conventional techniques (22, 25), using oligo(dT) or oligo(dT)-tailed pUC9 to prime cDNA synthesis, no clones containing long cDNA inserts were obtained. Therefore, a PCR-based method was used to generate clones containing the 3'-terminal region of the RNA. cDNA was prepared from the poly(A)-tailed CABYV RNA using the oligonucleotide 5'-GCC-AAGCT₁₈ (oligo 5) to prime cDNA synthesis. The cDNA was then subjected to PCR using oligo 5 and oligo 4 (equivalent to nucleotides 3396–3415, Fig. 1B). The resulting 2274-bp PCR product was digested with *Hind*III, which produced fragments of about 1800 and 500 bp. The two fragments were purified and the larger was cloned into *Hind*III-cleaved pBS(–) (Stratagene) and the smaller into *Hind*III/*Sma*I-cleaved pBS(–) to produce pCA36 and pCA6, respectively (Fig. 1B). The insert of each clone was completely sequenced on both strands using the universal direct and reverse primers and primers specific for internal insert sequences. Sequence data were also obtained for subclones of the pCA36 insert.

tein or degradation products thereof can be detected in association with partially purified virus particles (18, 28) and it has been suggested that the RT domain itself, which is subject to partial proteolytic degradation in the course of virus purification (18), is probably exposed on the virion surface where it may contain determinants important in virus-vector interactions.

Multiple sequence alignment of the RT domain of all luteoviruses reveals considerable homology (Fig. 3). The region immediately downstream of the coat protein termination codon is very rich in proline. This "proline hinge" (Fig. 3) may serve as a loose tether joining the coat protein moiety (presumably anchored in the virion capsid) to the rest of the RT domain. The rest of the RT domain can be divided into two parts: the C-terminal distal region in which there are a large number of positions absolutely conserved homology among all the sequenced luteoviruses (the "Luteovirus Homology Subdomain"; Fig. 3) and the C-terminal proximal region, which displays much less overall homology.

CABYV, like BWYV and PLRV, is efficiently transmitted by *M. persicae* but this aphid is not an efficient vector for BYDV(RPV), BYDV(MAV), and BYDV(PAV) (29). Evidently, comparisons between the CP and RT domain sequences of these two classes of virus could identify "*M. persicae*-specific" sequence motifs. This analysis reveals that such motifs are more abundant in the RT domain than in the CP. Thus, the aligned coat protein sequences of the aforesaid luteoviruses contain only two positions with the same amino acid in CABYV, BWYV, and PLRV but absent from the other luteoviruses (data not shown). The C-terminal portion of the RT domain, on the other hand, contains a high content of such vector-specific residues with, in particular, a region about 70 residues from the C-terminus (bracketed in Fig. 3) in which 17 of 44 residues are identical exclusively in CABYV, BWYV, and PLRV. It will be interesting to determine if this region is of particular importance in conferring specificity on the aphid transmission process.

ACKNOWLEDGMENTS

The authors thank Daniele Scheidecker for technical assistance and Olivier Poch for advice concerning sequence alignments.

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